

BAR Barbiturates strip CE

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For professional *in vitro* diagnostic use only

Barbiturates

A rapid test for the qualitative detection of Barbiturates in human urine.
ONE STEP

PRINCIPLE

The LINEAR BAR Barbiturates strip is an immunoassay based on the principle of competitive binding. Drugs that may be present in the urine specimen compete against the drug conjugate for binding sites on the antibody.

During testing, a urine specimen migrates upward by capillary action. Barbiturates, if present in the urine specimen below the 300 ng/ml, will not saturate the binding sites of the antibody in the test strip. The antibody coated particles will then be captured by immobilized Barbiturates-protein conjugate and a visible colored line will show up in the test line region. The colored line will not form in the test line region if the Barbiturates level exceeds the cut-off level because it will saturate all the binding sites of anti-Barbiturates antibodies.

A drug-positive urine specimen will not generate a colored line in the test line region because of drug competition, while a drug-negative urine specimen or a specimen containing a drug concentration less than the cut-off will generate a line in the test line region.

To serve as a procedural control, a colored line will always appear at the control line region indicating that proper volume of specimen has been added and membrane wicking has occurred.

REAGENT COMPOSITION

BAR Barbiturates test strip, contains mouse monoclonal anti-Barbiturates antibody-coupled particles and Barbiturates-protein conjugate. A goat antibody is employed in the control line system.

PACKAGING CONTENTS

REF 4415150 50 BAR Barbiturates test strips.

STORAGE AND STABILITY

Store at 2-30°C.

The test strip is stable through the expiration date printed on the sealed pouch. The test strip must remain in the sealed pouch until use. **DO NOT FREEZE.** Do not use beyond the expiration date.

SPECIMEN COLLECTION AND PREPARATION

The urine specimen must be collected in a clean and dry container. Urine collected at any time of the day may be used. Urine specimens exhibiting visible particles should be centrifuged, filtered, or allowed to settle to obtain clear specimen for testing.

Urine specimens may be stored at 2-8°C for up to 48 hours prior to testing. For long-term storage, specimens may be frozen and stored below -20°C. Frozen specimens should be thawed and mixed before testing.

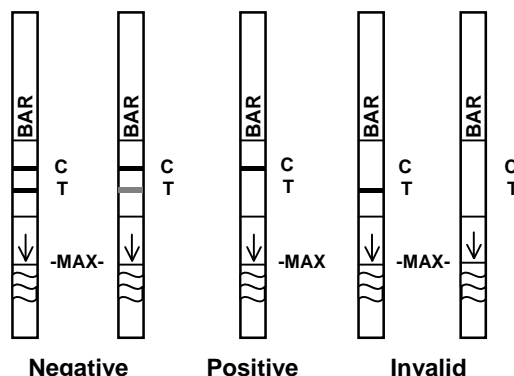
MATERIAL REQUIRED

- Timer.
- Specimen collection container.

PROCEDURE

Allow the test strip, urine specimen, and/or controls to reach room temperature (15-30°C) prior to testing.

1. Bring the pouch to room temperature before opening it. Remove the test strip from the sealed pouch and use it as soon as possible.
2. With arrows pointing toward the urine specimen, **immerse the test strip vertically in the urine specimen for at least 10-15 seconds.** Do not pass the maximum line (MAX) on the test strip when immersing the strip. See the illustration below.
3. Place the test strip on a non-absorbent flat surface, start the timer and wait for the red line(s) to appear. The result should be **read at 5 minutes.** Do not interpret the result after 15 minutes.



NEGATIVE: * **Two lines appear.** One colored line should be in the control region (C), and another apparent colored line should be in the test region (T). This negative result indicates that the Barbiturate concentration is below the detectable cut-off level.

* **NOTE:** The shade of red in the test line region (T) may vary, but it should be considered negative whenever there is even a faint colored line.

POSITIVE: **One colored line appears in the control line region (C).** No line appears in the test line region (T). This positive result indicates that the Barbiturate concentration exceeds the detectable cut-off level.

INVALID: **Control line fails to appear.** Insufficient specimen volume or incorrect procedural techniques are the most likely reasons for control line failure. Review the procedure and repeat the test using a new test strip. If the problem persists, discontinue using the lot immediately and contact your local distributor.

QUALITY CONTROL

A procedural control is included in the test. A red line appearing in the control region (C) is considered an internal procedural control. It confirms sufficient specimen volume, adequate membrane wicking and correct procedural technique. Control reagents are not supplied with this kit; however, it is recommended that positive and negative controls be tested as good laboratory testing practice to confirm the test procedure and to verify proper test performance.

CLINICAL SIGNIFICANCE

Barbiturates are central nervous system depressants. They are used therapeutically as sedatives, hypnotics, and anticonvulsants. Barbiturates are almost always taken orally as capsules or tablets. The effects resemble those of intoxication with alcohol. Chronic use of Barbiturates leads to tolerance and physical dependence. Short acting Barbiturates taken at 400 mg/day for 2-3 months produces a clinically significant degree of physical dependence. Withdrawal symptoms experienced during periods of drug abstinence can be severe enough to cause death. Only a small amount (less than 5%) of most Barbiturates are excreted unaltered in the urine. The detection period for the Barbiturates in the urine is 4-7 days.¹

The LINEAR BAR Barbiturates strip is a rapid urine-screening test that can be performed without the use of an instrument. The test utilizes a monoclonal antibody to selectively detect elevated levels of Barbiturates in urine. The LINEAR BAR Barbiturates strip yields a positive result when the Barbiturates in urine exceed the cut-off level 300 ng/mL.

Test to monitor therapeutic measures related to the study and control of detoxification treatments of drug of abuse and its effects in laboratory tests.

ANALYTICAL PERFORMANCE

A. Accuracy

The accuracy of the LINEAR BAR Barbiturates strip was compared and checked against a commercially available test with a threshold value of 300 ng/mL. 120 urine samples taken from volunteer test persons who claim to be non-consumers was examined under both tests. The results were 100% in agreement.

B. Reproducibility

The reproducibility of the LINEAR BAR Barbiturates strip was verified by blind tests performed at a four different locations. All 60 utilized samples with a BAR-concentration of 150 ng/mL yielded a negative result. All 60 samples with a BAR-concentration of 600 ng/mL yielded a positive result. No significant differences were observed between test results of the different evaluation sites.

C. Precision

Test precision was determined by blind tests with control solutions. Controls with a BAR-concentration of 150 ng/mL yield a negative result. Controls with a BAR-concentration of 600 ng/mL provide a positive result.

D. Specificity

The specificity of the LINEAR BAR Barbiturates strip was tested with the substances listed below, all of which can be found in a normal urine specimen. These substances were added to normal drug free urine.

The following compounds with a similar chemical structure yielded a positive result at the specified concentration:

COMPOUND	CONCENTRATION (ng/mL)
Secobarbital	300
Phenobarbital	300
Butalbital	3,000
Allobarbital	5,000
Alphenal	625
Amobarbital	600
Aprobarbital	600
Hexobarbital	>100,000
Butabarbital	75
Pentobarbital	300

All following listed compounds reacted negative up to a concentration of 100 µg/mL.

Acetamidophene	Guaiacol Glyceryl Ether
Acetone	Hemoglobin
Albumin	Imipramine
Amitriptyline	(+/-)-Isoproterenol
Ampicillin	Lidocaine
Aspartame	(+)-Naproxen
Aspirin	Oxalic Acid
Atropine	Penicillin-G
Benzocaine	Pheniramine
Bilirubin	Phenothiazine
Caffeine	Phenylethylamine
Chloroquine	Procaine
(+/-)-Chlorpheniramine	Quinidine
Chlorpheniramine	Ranitidine
Creatine	Riboflavin
Dexbrompheniramine	Sodium Chloride
Dextromethorphan	Sulindac
4-Dimethylaminoantipyrine	Thioridazine
Dopamine	Trifluoperazine
Erythromycin	Trimethobenzamide
Ethanol	Tyramine
Furosemide	Vitamin C
Glucose	

NOTES

1. The LINEAR BAR Barbiturates strip provides only a qualitative, preliminary analytical result. A secondary analytical method must be used to obtain a confirmed result. Gas chromatography/mass spectrometry (GC/MS) is the preferred confirmatory method.
2. Adulterants, such as bleach and/or alum, in urine specimens may produce erroneous results regardless of the analytical method used. If adulteration is suspected, the test should be repeated with another urine specimen.
3. A positive result indicates presence of the drug or its metabolites but does not indicate level of intoxication, administration route or concentration in urine.
4. A negative result may not necessarily indicate drug-free urine. Negative results can be obtained when drug is present but below the cut-off level of the test.
5. Test does not distinguish between drugs of abuse and certain medications.

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